

# **Lyoph-Ready Cesium Klentag C**

Amount: 4000 x 25 µl reactions up to 1 kb (equivalent to 200ul standard enzyme. Volume may be up to 2.5x higher)

Cat #: GF280

**Shipping conditions:** Ice Pack

Storage conditions: 4°C for 4 months or -20°C for 2 years with up to 10 freeze/thaw cycles

**Thermostability:** Retains at least 85% activity after 1 hour at 95°C

Expiration: On tube label

#### PRODUCT DESCRIPTION:

A lyoph-ready preparation of Cesium Klentaq C, a cold-sensitive mutant of Klentaq1 (5'-exonuclease deficient Taq polymerase with improved fidelity and thermostability). Due to its suppressed activity at low temperatures this enzyme is designed for hot start PCR performance. Since Omni Klentaq lacks 5' exonuclease activity, it is not suitable for TaqMan assays. 10x buffer composition is: 500 mM Tris-Cl pH 9.2, 160 mM ammonium sulfate, 0.25% Brij 58, and 35 mM magnesium chloride.

### TYPICAL PCR PROTOCOL for a 25 µl reaction:

Reagent	Volume	Final Concentration
10x Klentaq Mutant Reaction Buffer	2.5 μl	1x
dNTP mix (10 mM each)	0.5 μl	200 μM each
Left Primer	variable	200 nM
Right Primer	variable	200 nM
DNA template†	variable	0.1-100 ng
PCR Enhancer Cocktail (recommended)*	12.5 μl	1x
Cesium Klentaq C	0.05 – 0.25 μl **	
De-ionized distilled H2O	Adjust final volume to 25 μl	

<sup>†</sup> DNA amount depends mostly on genome size and target gene copy number.

#### **CYCLING CONDITIONS:**

1. Denaturing: 94° for 2-8 minutes for 1 cycle

2. Denaturing: 94° for 40-60 seconds

3. Annealing: 50°-68° depending on the specific Tm primers for 40-60 seconds

4. Extension: 68° for 2 min/kb target

5. Repeat steps 2-4 for 25-40 cycles

Please visit us on the web at www.klentaq.com for troubleshooting and detailed protocols.

## **REFERENCES:**

Kermekchiev, M.B., et al. (2003) Cold-sensitive mutants of Taq DNA polymerase provide a hot start for PCR. Nucl Acids Res. 31, 6139-6147.

Kermekchiev, M.B. et al. (2009) Mutants of Taq DNA polymerase resistant to PCR inhibitors allow DNA amplification from whole blood and crude soil samples. Nucl. Acids Res., 37 (5):e40 E pub.

<sup>\*</sup> For optimal performance, we recommend using one of our PCR Enhancer Cocktails (PEC-1, PEC-1GC, PEC-2, or PEC-2-GC) or 1.3 M Betaine, a general PCR enhancer.

<sup>\*\*</sup> To determine specific optimal enzyme concentration, we strongly recommend an enzyme titration test for each target. A good starting amount of the enzyme per 25  $\mu$ l reaction is 0.05  $\mu$ l. Targets larger than 1 kb may require more enzyme or may benefit from the use of an LA (Long Accurate) version of the polymerase.