Hot Start OmniTaq 2 LA

Amount: 125 μl (500 x 25 μl reactions)

Shipping conditions: Ambient **Storage conditions:** -20°C

Thermostability: Retains at least 85% activity after 1 hour at 95°C

Expiration: On tube label



PRODUCT DESCRIPTION:

Hot Start OmniTaq 2 LA is made with aptamer-based technology, enabling room temperature reaction set-up.

OmniTaq 2 is a mutant of Taq DNA polymerase. This enzyme provides 2-3x faster PCR and some inhibition-resistance. The aptamer binds to the polymerase at sub-cycling temperatures, inactivating the enzyme and preventing spurious amplification. The Long-Accurate feature allows for amplification of longer products with higher fidelity and accuracy. LA enzymes are not recommended for use with dUTP. 10x buffer composition is: 500 mM Tris-Cl pH 9.1, 160 mM ammonium sulfate, 0.25% Brij 58, and 25 mM magnesium chloride.

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TYPICAL PCR PROTOCOL for a 25 µl reaction:

Reagent	Volume	Final Concentration
10x Taq Mutant Reaction Buffer	2.5 μl	1x
dNTP mix (10 mM each)	0.5 μl	200 μM each
Left Primer	variable	200 nM
Right Primer	variable	200 nM
DNA template†	variable	Up to 1 ng/μl
PCR Enhancer Cocktail (optional)*	12.5 μ1	1x
Hot Start OmniTaq 2 LA	0.05 – 0.25 μl **	
De-ionized distilled H2O	Adjust final volume to 25 µl	

[†] DNA amount depends mostly on genome size and target gene copy number.

CYCLING CONDITIONS FOR PCR:

- 1. Initial Denaturing: 94° for 2-8 minutes recommended for crude samples containing 5-10% whole blood, plasma or serum.
- 2. Denaturing: 94° for 40-60 seconds
- 3. Annealing: 50°-68° depending on the specific Tm primers for 40-60 seconds
- 4. Extension: 68° for 1 min/kb target
- 5. Repeat steps 2-4 for 25-40 cycles

Please visit us on the web at www.klentaq.com for troubleshooting and detailed protocols.

REFERENCES:

Barnes, W. M., et al. (2021) A Single Amino Acid Change to Taq DNA Polymerase Enables Faster PCR, Reverse Transcription and Strand-Displacement. *Frontiers in Bioengineering and Biotechnology*. 8:553474. https://doi.org/10.3389/fbioe.2020.553474

^{*} When inhibition-resistance is needed, we recommend using one of our PCR Enhancer Cocktails (PEC-1, PEC-1GC, PEC-2, or PEC-2-GC) which are specially formulated for use with whole blood, serum or plasma or 1.3M Betaine, a gerneric PCR enhancer.

^{**} To determine specific optimal enzyme concentration, we strongly recommend an enzyme titration test for each target. A good starting amount of the enzyme per 25 μ l reaction is 0.05 μ l for purified DNA templates and 0.25 μ l for crude samples containing 5-10% whole blood, plasma or serum. Targets larger than 1 kb may require more enzyme.