

# **Lyoph-Ready Klentaq1** Cat #: GF100

Amount: 4000 x 25 µl reactions up to 1 kb (equivalent to 200ul standard Klentaq1. Volume may be up to 2.5x higher)

**Shipping conditions:** Ice Pack

Storage conditions: 4°C for 4 months or -20°C for 2 years with up to 10 freeze/thaw cycles

Thermostability: Retains at least 85% activity after 1 hour at 95°C

**Expiration**: On tube label

#### PRODUCT DESCRIPTION:

A lyoph-ready preparation of Klentaq1 suitable for lyophilization, is a 5'-exonuclease deficient Taq polymerase (an N-terminal deletion of Taq) with improved fidelity and thermostability. 10x buffer composition is: 500 mM Tris-Cl pH 9.2, 160 mM ammonium sulfate, 0.5% Brij 58, and 35 mM magnesium chloride. We also offer (upon request) 10x buffer at pH 7.9 for better fidelity.

## TYPICAL PCR PROTOCOL for a 25 ul reaction:

Reagent	Volume	<b>Final Concentration</b>
10x Klentaq1 reaction buffer	2.5 µl	1x
DNTP mix (10 mM)	0.5 ul	200 uM each
Left Primer	variable	200 nM
Right Primer	variable	200 nM
DNA template†	variable	0.1-100 ng
PCR Enhancer Cocktail (recommended)*	12.5 μl	1x
Klentaq1**	0.05 – 0.25 μl **	
De-ionized distilled H <sub>2</sub> O	Adjust final volume to 25 ul	-

<sup>†</sup> DNA amount depends mostly on genome size and target gene copy number.

Note: Enzyme amount per reaction should be adjusted based on final enzyme concentration.

## CYCLING CONDITIONS

1. Denaturing: 94° for 2 minutes for 1 cycle

2. Denaturing: 94° for 40-60 seconds

3. Annealing: 50°-68° depending on the specific primers (5° less than Tm) for 40-60 seconds

4. Extension: 68° for 2 min/kb target

5. Repeat steps 2-4 for 25-40 cycles

Please visit us on the web at www.klentaq.com for troubleshooting and detailed protocols.

#### **REFERENCES:**

Barnes, W.M. (1994) PCR amplification of up to 35 kb DNA with high fidelity and high yield from bacteriophage templates, PNAS 91, 2216-2220.

U.S. Patent No. 5,436,149

<sup>\*</sup> For optimal performance, we recommend using one of our PCR Enhancer Cocktails (PEC-1, PEC-1GC, PEC-2, or PEC-2-GC) or 1.3 M Betaine, a general PCR enhancer.

<sup>\*\*</sup> To determine specific optimal enzyme concentration, we strongly recommend an enzyme titration test for each target. A good starting amount of the enzyme per 25 µl reaction is 0.05 µl. Targets larger than 1 kb may require more enzyme or may benefit from the LA (Long-Accurate) version of the polymerase.